



# Sports nutrology and gut microbiota: a systematic review

Lucas Vasconcelos Lima Diniz<sup>1\*</sup>

<sup>1</sup> Nutroleve Medicina e Treinamento, Macapá, Amapá, Brazil.

\*Corresponding Author: Dr. Lucas Vasconcelos Lima Diniz.  
Nutroleve Medicina e Treinamento, Macapá, Amapá, Brazil.  
E-mail: vasconceloslucasilmadiniz@gmail.com

DOI: <https://doi.org/10.54448/ijn23109>

Received: 10-18-2022; Revised: 01-10-2022; Accepted: 01-23-2023; Published: 01-25-2023; IJN-id: e23109

## Abstract

**Introduction:** Many of the established positive health benefits of exercise have been documented by historic discoveries in the field of exercise physiology. Regular physical training associated with nutritional health has broad health benefits for the gut microbiota, acting positively on almost all organ systems of the body.

**Objective:** It was to analyze the main metabolic pathways modulated by nutrients, gut microbiota, and physical exercise in muscle regeneration and sports performance. **Methods:** The present study followed a systematic review model (PRISMA). The literary search process was carried out from July to September 2022 and was developed based on Scopus, PubMed, Science Direct, Scielo, and Google Scholar, with scientific articles from 2004 to 2022. The low quality of evidence was attributed to case reports, editorials, and brief communications, according to the GRADE instrument. The risk of bias was analyzed according to the Cochrane instrument.

**Results and Conclusion:** We found 132 studies that underwent eligibility analysis, and then 31 of the 52 total studies were selected for this systematic review. According to the GRADE instrument, most studies showed homogeneity in their results, with  $I^2 = 98.9\% > 50\%$ . The Funnel Plot showed a symmetrical behavior, not suggesting a significant risk of bias in studies with a smaller sample size. A healthy gut microbiota and a positive interaction with the immune system, promoted by diligent nutrological care, can be crucial for the muscle-gut axis and can influence the maintenance of muscle mass and its functionality in athletes. However, dysbiosis resulting from a negative interaction with the immune system can influence muscle wasting disorders. These changes can promote systemic inflammation, with overproduction of the pro-

inflammatory cytokines TNF- $\alpha$ , IL-1 $\beta$ , and IL-6. Future studies should clarify whether gut microbiota dysbiosis and nutrient depletion are pathophysiologically associated with muscle wasting disorders and whether exercise can positively influence this supposed gut-muscle axis.

**Keywords:** Nutrients. Nutrology. Gut microbiota. Sports. Metabolism. Skeletal muscle.

## Introduction

Many of the established positive health benefits of exercise have been documented by historic discoveries in the field of exercise physiology. These investigations generally assess performance thresholds or exercise-induced health benefits [1]. Thus, several important findings were informed by the study of athletes. Recent progress has been made concerning gut microbiota (GM), regenerative nutrition, and skeletal muscle metabolism [1-3].

In this context, regular physical training associated with nutritional health has broad benefits for the health of the GM, acting positively on almost all organ systems of the body [4]. The mysteries of human physiology and the adaptive response to acute and chronic physical training have been largely elucidated through exercise science. Thus, exercise physiologists have studied the physiological response to physical activity and sports [5,6].

In the context of the triad physical exercise, nutrition, and intestinal microbiota for the process of muscle regeneration, adult stem cells stand out as gut stem cells at the base (crypts) of the intestine and muscle stem cells outside the sarcolemma adjacent to the muscle basement membrane [7-9]. The tissue

niche is also able to influence adult stem cells metabolism. Tissue stem cell metabolism has focused on central carbon metabolism, that is, the generation of metabolic building blocks via glycolysis, oxidative phosphorylation, or the pentose phosphate pathway.

Adult tissue stem cells mediate tissue and organ homeostasis and regeneration, making decisions about whether to remain quiescent, proliferate, or differentiate into mature cell types. These decisions are directly integrated with the body's energy balance and nutritional status. Metabolic by-products and substrates that regulate epigenetically and signaling pathways are considered to have an instructive role, rather than an observer, in the regulation of cell fate decisions [9].

In this sense, it is suspected that the quiescent state of stem cells is characterized by an inherently glycolytic metabolism, followed by a transition to favor mitochondrial oxidative phosphorylation during differentiation [10-13]. However, increasing evidence suggests that metabolism during quiescence, activation, and differentiation may vary between tissues, integrating signaling cues and metabolic inputs from the niche and the organism as a whole, mainly by signaling nutrients and the gut microbiota. In this scenario, metabolomics provides information on cellular pathways, observing substrates and metabolic products through different pathways [14,15]. Along with transcriptomics and proteomics analysis, it is observed that metabolism can affect cell fate (and vice versa) [16].

In the sports practice scenario, both physical exercise and nutrients are modulators of the GM composition, increasing biodiversity and beneficial metabolic functions [17]. Overtraining is associated with IM dysbiosis, promoting inflammation and negative metabolic consequences [17,18].

Furthermore, GM can influence the pathophysiology of several distant organs, including skeletal muscle [19,20]. The gut-muscle axis can regulate muscle protein deposition and muscle function [17]. In older individuals, this axis may be involved in the pathogenesis of muscle wasting disorders through multiple mechanisms, involving transduction of pro-anabolic stimuli from dietary nutrients, modulation of inflammation, and insulin sensitivity [21].

In this sense, the immune system plays a key role in these processes, being influenced by the composition of the microbiome and at the same time contributing to the formation of microbial communities. In this sense, exercise is considered one of the main environmental factors that possibly influence the composition of GM [21,22].

Therefore, the present systematic review study analyzed the main metabolic pathways modulated by nutrients, gut microbiota, and physical exercise in muscle regeneration and sports performance.

## Methods

### Study Design

The present study followed a concise systematic review model, following the rules of systematic review - PRISMA (Transparent reporting of systematic review and metaanalysis- [www.prisma-statement.org/](http://www.prisma-statement.org/))

### Search Strategy and Search Sources

The literary search process was carried out from July to September 2022 and was developed based on Scopus, PubMed, Science Direct, Scielo, and Google Scholar, with scientific articles from 2004 to 2022, using the descriptors (MeSH Terms): "Nutrients. Nutriology. Intestinal microbiota. Sports. Metabolism. Skeletal muscle", and using the Booleans "and" between the MeSH terms and "or" between the historical findings.

### Study Quality and Risk of Bias

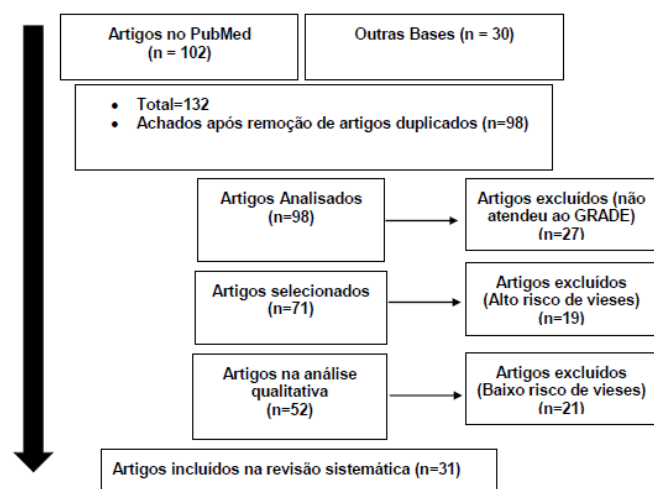
Quality was rated as high, moderate, low, or very low for risk of bias, clarity of comparisons, precision, and consistency of analyses. The most evident highlight was for systematic review articles or meta-analysis of randomized clinical trials, followed by randomized clinical trials. The low quality of evidence was attributed to case reports, editorials, and brief communications, according to the GRADE instrument. The risk of bias was analyzed according to the Cochrane instrument through the analysis of the Funnel Plot (Cohen test (d)).

## RESULTS AND DISCUSSION

### Summary of Literary Findings

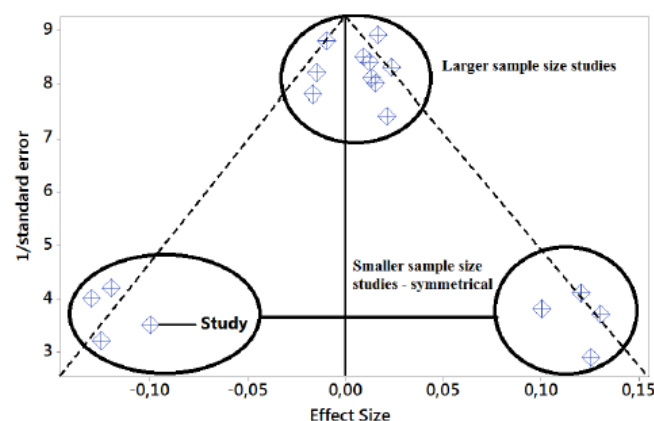
It was found 132 studies that underwent eligibility analysis and, then, 31 of the 52 total studies were selected for the present systematic review (Figure 1), considering in the first instance the level of scientific evidence of studies in study type such as metaanalysis, consensus, randomized clinical trial, prospective and observational. The biases did not compromise the scientific basis of the studies. According to the GRADE instrument, most studies showed homogeneity in their results, with  $R^2 = 98.9\% > 50\%$ .

**Figure 1.** Flowchart showing the article selection process.



**Figure 2** presents the results of the risk of bias of the studies using the Funnel Plot, showing the calculation of the Effect Size (Magnitude of the difference) using the Cohen Test (d). Precision (sample size) was determined indirectly by the inverse of the standard error (1/Standard Error). This graph had a symmetrical behavior, not suggesting a significant risk of bias, both between studies with a small sample size (lower precision) that are shown at the bottom of the graph and in studies with a large sample size that are presented in the upper region.

**Figure 2.** The symmetrical funnel plot suggests no risk of bias between the small sample size studies that are shown at the bottom of the plot. High confidence and high recommendation studies are shown above the graph (NTotal=31 studies evaluated in full in the systematic review).



## Metabolic Pathways and Muscle Regeneration

Physical activity, endogenous metabolites, and dietary nutrients can directly influence epigenetic

enzymes. Epigenetic modifications in DNA and histone proteins alter the fate of the cell by controlling chromatin accessibility and downstream gene expression patterns [16].

Thus, many substrates and cofactors for chromatin-modifying enzymes are derived from metabolic pathways involving the tricarboxylic acid cycle, the methionine cycle, the folate cycle, glycolysis,  $\beta$ -oxidation, and the hexosamine pathway. These metabolites can serve as activators or inhibitors of epigenetic writers, such as proteins containing the Jumonji C domain (JmjC), DNA methyltransferases (DNMTs), histone acetyltransferases (HATs), ten-eleven DNA translocase demethylases (TETs) and histone deacetylases (HDACs). In this sense, metabolites can influence nutrient detection signaling pathways [16].

Thus, the mechanistic target of the rapamycin complex 1 (mTORC1) can be activated by growth factor-induced signaling only when the amino acids arginine and leucine, as well as the cofactor S-adenosyl methionine (SAM), are detected within the cell. Furthermore, the energy balance communicated through the cellular AMP/ADPATP ratio can be detected by AMP-activated protein kinase (AMPK). In addition, transcription factors can be directly regulated by metabolites, for example, the tryptophan kynurenine metabolite is an endogenous agonist of the aryl hydrocarbon receptor and  $\alpha$ -ketoglutarate ( $\alpha$ -KG) binds and activates IKK $\beta$  and initiates IKK $\beta$  signaling. NF $\kappa$ B [16].

In this scenario, dietary manipulations and metabolites can affect tissue stem cell fate decisions, as highlighted in the small intestine (intestinal stem cells (LGR5+)), hematopoietic system (hematopoietic stem cells (HSCs), liver, muscle (muscle stem cells/satellite cells) and hair follicles (hair follicle stem cells (HFSCs)). For example, in HFSCs, mitochondrial pyruvate carrier 1 (MPC1) and lactate dehydrogenase (LDHA) regulate the balance between telogen and anagen during the hair cycle. In LGR5+, 3hydroxy-3-methylglutaryl-CoA synthase (Hmgcs2) is highly expressed while MPC1/2 is expressed at low levels. The ketogenic or glucose-rich diet regulates the balance of selfrenewal of LGR5+ Self-renewal and differentiation of HSC can be regulated by manipulating vitamin C, A, or D levels and valine restriction [16].

Regarding muscle regeneration, a diet rich in nicotinamide riboside can increase muscle stem cell numbers and function in a histone deacetylase (SIRT1) dependent manner. Muscle stem cells, called satellite cells, are responsible for maintaining adult muscle mass and repairing it after injury. Several studies have

demonstrated how changes in innate metabolism interfere with the differentiation of satellite stem cells into mature myocytes [23]. For example, mapping a single cell with histone acetylation showed that acetylation levels tend to be low in quiescent cells.

In this context, one study found that isolated quiescent muscle stem cells express fatty acid oxidation enzymes/transporters, however, as they exit quiescence and enter the cell cycle for proliferation, a metabolic transition occurs to favor glycolysis [24]. In this sense, SIRT1 is a target of increased glycolysis. SIRT1 represses the maturity expression of specific skeletal muscle genes as well as genes involved in mitochondrial biogenesis.

Advanced glycolysis depletes NAD<sup>+</sup>, an essential metabolic cofactor of SIRT1, reducing SIRT1 activity and promoting downstream activation of these mature muscle-specific genes and differentiation [25].

### Physical Activity and Gut Microbiota

In a case-control study, microbial diversity was much greater in a group of professional players than in age, sex, and body-size-matched controls who do not play sports [26]. Recently, metagenomic analyzes of fecal samples from the same groups highlighted that athletes had a different microbial composition also from a functional point of view, with an increased microbial representation of genes involved in carbohydrate and amino acid metabolism, and fatty acid chain production [27].

In another study, the mean abundance of taxa involved in energy and carbohydrate metabolism, including *Prevotella* and *Methano-brevibacter smithii*, was significantly higher in professional than amateur cyclists and was correlated with training frequency [28]. However, these studies were unable to fully separate the contribution of exercise and diet in determining the different compositions of the microbiota in different groups, as participants followed a wide range of dietary regimens.

Also, the intensity of training is also important. Light exercise programs induce only slight changes in the composition of the gut microbiota in sedentary individuals [29]. Therefore, findings from studies performed on athletes should not be automatically transferred to all individuals who perform a non-competitive exercise.

According to three different studies, fecal microbiota biodiversity is correlated with cardiorespiratory fitness in adult individuals. However, in one of these studies, performed on 71

premenopausal Finnish women, this relationship was mediated by body composition [29-31]. Another study, performed on 19 active and 21 sedentary women aged ≤40 years, confirmed that the microbiome abundance of various bacterial taxa was significantly correlated with body fat or lean mass percentage [32]. Thus, the possible association between exercise and microbiota should be further investigated, taking into account possible confounding factors, such as dietary habits, nutrient intake, and body composition parameters [33].

The influence of body composition on the microbiota was also emphasized by the findings of an intervention study, where two groups of sedentary individuals, one lean and one obese, underwent a 6-week structured exercise program, followed by a washout period of 6 weeks. 6 weeks [34]. After exercise training, lean and obese participants experienced a change in gut microbiota composition, but the overall representation of species with known anti-inflammatory properties and the microbiome's ability to produce short-chain fatty acids (SCFA) was greater in lean subjects, highlighting a body mass index (BMI) dependent response to training. However, all changes reverted to the baseline state after the washout period [34].

Furthermore, in healthy, exercised young males undergoing a period of forced inactivity, cessation of the exercise was associated with changes in gastrointestinal physiology (i.e., reduced bowel movements and increased stool consistency) before changes in gastrointestinal GM composition and function could be detected [30-32]. These circumstances suggest that the microbiome is resilient to acute changes in exercise habits and that exercise maintenance is necessary to induce lasting changes in the gut microbiome.

In this aspect, changes in the composition of the gut microbiome induced by exercise can exert beneficial effects throughout the body, modulating pathological processes. For example, changes in the microbiota induced by exercise can attenuate the clinical course and evolution of experimental models of myocardial infarction or chemically induced colitis, mainly by modulating the inflammatory response [35]. The main mediators in these processes may be SCFAs, particularly butyrate, whose production by the gut microbiota has been shown to increase after exercise in humans [34].

### Nutrients and Gut microbiota In Muscle Pathophysiology

Several research groups have independently hypothesized that the composition of the gut microbiota may influence the onset of sarcopenia, ie the loss of

muscle mass and function that occurs with aging [17,19,30,31]. In this sense, a dysbiotic GM can reduce the bioavailability of dietary proteins and particularly of some amino acids, such as tryptophan, involved in modulating inflammation and promoting muscle protein synthesis [36-40]. Gut bacteria are also involved in the synthesis of many vitamins, including folate, vitamin B12, and riboflavin, exerting various beneficial and pro-anabolic effects on skeletal muscle cells, ranging from amino acid biosynthesis to neutralizing oxidative stress during exercise [41].

Furthermore, a healthy GM can effectively transform some dietary nutrients into metabolic mediators that, once absorbed into the systemic circulation, can exert beneficial effects on inflammation, insulin sensitivity, anabolism, and antioxidant capacity. Polyphenols, including resveratrol, and ellagitannins contained in pomegranates and berries represent the most relevant examples of nutrients that, after microbial metabolism, enter the systemic circulation and exert beneficial effects on the muscle [42,43]. Resistance training appears to increase the bioavailability of dietary polyphenols, likely through their beneficial modulations of GM [44].

In addition, age-related changes in GM composition, occurring regardless of the level of physical training, may promote intestinal mucosal dysfunction, with increased permeability. This phenomenon can result in the systemic absorption of microbial byproducts and toxins, including LPS. In skeletal muscle cells, circulating LPS may contribute to activating Toll-Like Receptors (TLR) 4 and 5, promoting the activation of the NF- $\kappa$ B pathway, with reduced insulin sensitivity, increased protein catabolism, and inflammatory cytokine production [17]. In aging humans, TLR4 activation is associated with metabolic endotoxemia, decreased insulin sensitivity, and reduced quadriceps muscle strength and volume.

In this context, the most studied mechanism involved in the gut modulation of GM by physical exercise is the bacterial production of metabolic mediators, including bile acids and SCFA [18]. A healthy GM can produce secondary bile acids, which are wellknown activators of the farnesoid X receptor stimulating myocyte anabolism. SCFA and particularly butyrate, are generally synthesized by a large number of intestinal bacteria, including *Faecalibacterium*, *Butyricimonas*, and *Succinivibrio*, highly represented in healthy individuals, but reduced in older individuals. These mediators have several beneficial metabolic activities, influencing skeletal muscle protein deposition by modulating the systemic anabolic/catabolic balance

[18].

Also, dietary polyphenols exert several beneficial effects on sports performance, demonstrated in vivo and in human studies. The health-related mechanisms of polyphenols mainly concern the modulation of mitochondrial biogenesis and stimulation of stress-related enzymes or transcription factors, as well as a nutritional deficiency, which regulates gene expression of essential antioxidant proteins (SOD, Catalase, Glutathione system, etc.). They have also been shown to modulate inflammatory processes and the immune system response (Th1/Th2 balance). Furthermore, some polyphenols favor vascular regulation and endothelial function in humans, increasing endothelial synthesis of nitric oxide [45].

Overall, these mechanisms promote athletic performance by improving cardiometabolic functions, reducing recovery times and post-exercise pain, maintaining a low degree of oxidative stress, and preventing dysregulated inflammatory processes. Thus, polyphenols are able, through their interaction with the intestinal microbiota, to favor the proliferation of bacterial genera of great importance for metabolic and cognitive functions, such as *Akkermansia*, *Lactobacilli*, and *Bifidobacteria*. The microbiota, on the other hand, metabolizes polyphenols in the colon to produce small bioactive molecules that exert epigenetic mechanisms in biochemical pathways modulating gene expression. Polyphenols have multiple biological effects, and future exercise studies should be designed appropriately and specifically to determine the physiological interactions between exercise and the selected supplement, rather than just considering performance [46].

### Gut-Muscle Axis

Intestinal muscle communication in human pathophysiology can be bidirectional, with GM making a true weave between epigenetic factors, physical exercise, and skeletal muscle [17].

In healthy individuals who regularly perform physical activities, a homeostatic balance between GM and skeletal muscle is present, promoting a healthy GM composition. However, this balance between GM and skeletal muscle can be disrupted by a sedentary lifestyle or excessive exercise, resulting in GM dysbiosis. Other factors that promote dysbioses, such as medications or acute illness, may also be associated with reduced muscle mass and function. Dysbiosis influences intestinal permeability, systemic inflammation, anabolism, and nutrient availability. All these mechanisms are involved in muscle physiology and represent the substrates of the gut-muscle axis [17].

Thus, the gut-muscle axis may be bidirectional, with GM influencing muscle and exercise contributing to shaping the composition of the microbiota. The intensity and frequency of exercise can be of great importance in determining the prevailing axis and its pathophysiological consequences.

Besides, a healthy GM plays a key role in shaping the local and systemic immune response to intestinal bacteria throughout life, favoring the maintenance of tolerance towards commensal antigens and activation against pathogen antigens [41]. The gut with microbiological dysbiosis favors the loss of immune tolerance to commensals, the deficiency of epithelial barrier function, and an imbalance in the activation of antiinflammatory Treg, lymphocytes, and pro-inflammatory Th17. These phenomena may contribute to the onset of inflammatory and autoimmune diseases, including inflammatory bowel disease, type 1 diabetes, and multiple sclerosis [47,48].

Furthermore, the immune system is indeed capable of influencing the composition of GM at various levels. Both innate and adaptive immunity are involved. Possible mechanisms include the production of antimicrobial peptides from intestinal cells, mucus secretion, activation of immunoglobulin A (IgA), similar to TLR receptor activation, lymphocyte transfer, and differentiation, and the presence of natural T cells (iNKT) [42,49].

Thus, whatever mediators are involved, the balance between the immune system and GM can be strongly influenced by epigenetic factors. Positive modulators of GM composition, including regular exercise, can induce a beneficial balance with the immune system, aiding in the maintenance of health [50-52].

## Conclusion

It was concluded that a healthy gut microbiota and a positive interaction with the immune system, promoted by diligent nutrological care, can be crucial for the muscle-gut axis, and can influence the maintenance of muscle mass and its functionality in athletes.

However, dysbiosis resulting from a negative interaction with the immune system can influence muscle wasting disorders. These changes can promote systemic inflammation, with overproduction of the pro-inflammatory cytokines TNF- $\alpha$ , IL-1 $\beta$ , and IL-6. Future studies should clarify whether gut microbiota dysbiosis and nutrient depletion are pathophysiologically associated with muscle wasting disorders and whether exercise can positively influence this supposed gut-

muscle axis.

## Acknowledgement

Not applicable.

## Funding

Not applicable.

## Ethics approval

Not applicable.

## Informed consent

Not applicable.

## Data sharing statement

No additional data are available.

## Conflict of interest

The authors declare no conflict of interest.

## Similarity check

It was applied by Ithenticate@.

## About the license

© The author(s) 2023. The text of this article is open access and licensed under a Creative Commons Attribution 4.0 International License.

## References

1. Skorski S, Mujika I, Bosquet L, Meeusen R, Coutts AJ, Meyer T. The Temporal Relationship Between Exercise, Recovery Processes, and Changes in Performance. *Int J Sports Physiol Perform.* 2019;14(8):1015-1021. doi:10.1123/ijsspp.2018-0668.
2. Tobin MJ. Why Physiology Is Critical to the Practice of Medicine: A 40-year Personal Perspective. *Clin Chest Med.* 2019;40(2):243-257. doi:10.1016/j.ccm.2019.02.012.
3. Foster C, Rodriguez-Marroyo JA, de Koning JJ. Monitoring Training Loads: The Past, the Present, and the Future. *Int J Sports Physiol Perform.* 2017;12(Suppl 2):S22-S28. doi:10.1123/ijsspp.2016-0388.
4. Margaritelis NV, Paschalis V, Theodorou AA, Kyparos A, Nikolaidis MG. Redox basis of exercise physiology. *Redox Biol.* 2020;35:101499. doi:10.1016/j.redox.2020.101499.
5. Rueggsegger GN, Booth FW. Health Benefits of Exercise. *Cold Spring Harb Perspect Med.* 2018 Jul 2;8(7). pii: a029694.
6. Cheng AJ, Yamada T, Rassier DE, Andersson DC,

- Westerblad H, Lanner JT. Reactive oxygen/nitrogen species and contractile function in skeletal muscle during fatigue and recovery. *J Physiol*. 2016 Sep 15;594(18):5149-60.
7. Ferraro, F. et al. Adult stem cells and their niches. *Adv. Exp. Med. Biol.* 2010, 695, 155–168
8. Blanpain, C. et al. Self-renewal, multipotency, and the existence of two cell populations within an epithelial stem cell niche. *Cell*. 2004, 118, 635–648.
9. Chacón-Martínez CA et al. Hair follicle stem cell cultures reveal selforganizing plasticity of stem cells and their progeny. *EMBO J*. 2017, 36, 151– 164.
10. Rodríguez-Colman, M.J. et al. Interplay between metabolic identities in the intestinal crypt supports stem cell function. *Nature* 2017, 543, 424.
11. Snoeck, H.W. Mitochondrial regulation of hematopoietic stem cells. *Curr. Opin. Cell Biol.* 2017, 49, 91–98.
12. Zheng, X. et al. Metabolic reprogramming during neuronal differentiation from aerobic glycolysis to neuronal oxidative phosphorylation. 2016, *Elife* 5, e13374.
13. Flores, A. et al. Lactate dehydrogenase activity drives hair follicle stem cell activation. *Nat. Cell Biol.* 2017, 19, 1017–1026
14. Rinschen MM. et al. Identification of bioactive metabolites using activity metabolomics. *Nat. Rev. Mol. Cell Biol.* 2019, 20, 353–367.
15. Agathocleous, M. et al. Ascorbate regulates haematopoietic stem cell function and leukaemogenesis. *Nature* 2017, 549, 476–481.
16. Shapira SN, Christofk HR. Metabolic Regulation of Tissue Stem Cells. *Trends Cell Biol.* 2020 Jul;30(7):566-576. doi: 10.1016/j.tcb.2020.04.004. Epub 2020 Apr 28. PMID: 32359707.
17. Ticinesi A, Lauretani F, Tana C, Nouvenne A, Ridolo E, Meschi T. Exercise and immune system as modulators of intestinal microbiome: implications for the gut-muscle axis hypothesis. *Exerc Immunol Rev.* 2019;25:84-95. PMID: 30753131.
18. Sorrenti, V.; Caudullo, G.; Lucignano, F.; Fortinguerra, S.; Zusso, M.; Giusti, P.; Buriani, A. Personalized sports nutrition: Role of nutrients in athletic performance. In *Sports, Exercise, and Nutritional Genomics*; Debmalya Barh, I.I.A., Ed.; Academic Press: Cambridge, MA, USA, 2019; pp. 411–431.
19. Kawabata, K.; Yoshioka, Y.; Terao, J. Role of Intestinal Microbiota in the Bioavailability and Physiological Functions of Dietary Polyphenols. *Molecules* 2019, 24, 370.
20. Schmidt TSB, Raes J, Bork P. The human gut microbiome: from association to modulation. *Cell*. 2018, 172: 1198-1215.
21. Bermon S, Petriz B, Kajeniene A, Prestes J, Castell L, Franco OL. The microbiota: an exercise immunology perspective. *Exerc Immunol Rev* 2015, 21: 70-79.
22. Codella R, Luzi L, Terruzzi I. Exercise has the guts: how physical activity may positively modulate gut microbiota in chronic and immune-based diseases. *Dig Liver Dis* 2018, 50: 331-341.
23. Ryall, J.G. and Lynch, G.S. The molecular signature of muscle stem cells is driven by nutrient availability and innate cell metabolism. *Curr. Opin. Clin. Nutr. Metab. Care*, 2018, 21, 240–245.
24. Machado, L. et al. In situ fixation redefines quiescence and early activation of skeletal muscle stem cells. *Cell Rep.* 2017, 21, 1982–1993.
25. Ryall, J.G. et al. The NAD<sup>+</sup>-dependent SIRT1 deacetylase translates a metabolic switch into regulatory epigenetics in skeletal muscle stem cells. *Cell Stem Cell*. 2015, 16, 171–183.
26. Batacan RB, Fenning AS, Dalbo VJ, Scanlan AT, Duncan MJ, Moore RJ, Stanley D. A gut reaction: the combined influence of exercise and diet on gastrointestinal microbiota in rats. *J Appl Microbiol* 2017, 122: 1627-1638.
27. Barton W, Penney NC, Cronin O, Garcia-Perez I, Molloy MG, Holmes E, Shanahan F, Cotter PD, O'Sullivan O. The microbiome of professional athletes differs from that of more sedentary subjects in composition and particularly at the functional metabolic level. *Gut* 2018, 67: 625-633.
28. Cronin O, Barton W, Skuse P, Penney NC, Garcia-Perez I, Murphy EF, Woods T, Nugent H, Fanning A, Melgar S, Falvey EC, Holmes E. Cotter PD, O'Sullivan O, Molloy MG, Shanahan F. A prospective metagenomics and metabolomics analysis of the impact of exercise and/or whey protein supplementation on the gut microbiome of sedentary adults. *mSystems* 2018, 3: e00044-18.
29. Durk RP, Castillo E, Marquez-Megana L, Grosicki GJ, Bolter ND, Lee CM, Bagley JR. Gut microbiota composition is related to cardiorespiratory fitness in healthy young adults. *Int J Sport Nutr Exerc Metab* ahead of print Jun 10, 2018. Doi: 10.1123/ijsnem.2018-0024.
30. Sket R, Debevec T, Kublik S, Schlöter M, Schoeller A, Murovec B, Vogel Mikus K, Makuc D, Pecnik K, Plavec J, Mekjavic IB, Eiken O, Prevorsek Z, Stres B. Intestinal metagenomes and metabolomes in healthy young males: inactivity and hypoxia generated negative physiological symptoms precede microbial dysbiosis. *Front Physiol* 2018, 9: 198.
31. Sket R, Treichel N, Debevec T, Eiken O, Mekjavic I, Schlöter M, Vital M, Chandler J, Tiedje JM, Murovec B, Prevorsek Z, Stres B. Hypoxia and inactivity related physiological changes (constipation, inflammation) are not reflected at the level of gut metabolites and butyrate producing microbial community: the PlanHab Study. *Front Physiol* 2017, 8: 250.
32. Sket R, Treichel N, Kublik S, Debevec T, Eiken O,

- Mekjavic I, Schlöter M, Vital M, Chandler J, Tiedje JM, Murovec B, Prevorsek Z, Linar M, Stres B. Hypoxia and inactivity related physiological changes precede or take place in absence of significant rearrangements in bacterial community structure: the PlanHab randomized trial pilot study. *PLoS One* 2017;12: e0188556.
33. Karl JP, Margolis LM, Madslie EH, Murphy NE, Castellani JW, Gundersen Y, Hoke AV, Levangie MW, Kumar R, Chakraborty N, Gautam A, Hammamieh R, Martini S, Montain SJ, Pasiakos SM. Changes in intestinal microbiota composition and metabolism coincide with increased intestinal permeability in young adults under prolonged physiological stress. *Am J Physiol Gastrointest Liver Physiol* 2017, 312: G559-G571.
  34. Zhao X, Zhang Z, Hu B, Huang W, Yuan C, Zou L. Response of gut microbiota to metabolite changes induced by endurance exercise. *Front Microbiol* 2018, 9: 765.
  35. De Sire R, Rizzatti G, Ingravalle F, Pizzoferrato M, Petito V, Lopetuso L, Graziani C, de Sire A, Mentella MC, Mele MC, Gasbarrini A, Scadaferri F. Skeletal muscle-gut axis: emerging mechanisms of sarcopenia for intestinal and extra-intestinal diseases. *Minerva Gastroenterol Dietol* 2018, 64: 351-362.
  36. Siddhart J, Chakrabarti A, Pannérec A, Karaz S, Morin-Rivron D, Masoodi M, Feige JN, Parkinson SJ. Aging and sarcopenia associate with specific interactions between gut microbes, serum biomarkers and host physiology in rats. *Aging* 2017, 9: 1698- 1720.
  37. Sung MM, Byrne NJ, Robertson IM, Kim TT, Samokhvalov V, Levasseur J, Soltys CL, Fung D, Tyreman N, Denou E, Jones KE, Seubert JM, Schertzer JD, Dyck JRB. Resveratrol improves exercise performance and skeletal muscle oxidative capacity in heart failure. *Am J Physiol Heart Circ Physiol* 2017, 312: H842-H853.
  38. Pereira-Cano G, Polyviou T, Ludwig IA, Nastase AM, Moreno-Rojas JM, Garcia AL, Malkova D, Crozier A. Bioavailability of orange juice (poly)phenols: the impact of short-term cessation of training by male endurance athletes. *Am J Clin Nutr* 2017, 106: 791-800.
  39. Cerdà B, Perez M, Perez-Santiago JD, Tornero-Aguilera JF, Gonzalez-Soltero R, Larrosa M. Gut microbiota modification: another piece in the puzzle of the benefits of physical exercise in health? *Front Physiol* 2016, 7: 51.
  40. Ticinesi A, Lauretani F, Milani C, Nouvenne A, Tana C, Del Rio D, Maggio M, Ventura M, Meschi T. Aging gut microbiota at the cross-road between nutrition, physical frailty and sarcopenia: is there a gut-muscle axis? *Nutrients* 2017, 9: E1303.
  41. Gensollen T, Iyer SS, Kasper DL, Blumberg RS. How colonization by microbiota in early life shapes the immune system. *Science* 2016, 352: 539544.
  42. Hooper LV, Littman DR, Macpherson AJ. Interactions between the microbiota and the immune system. *Science* 2012, 336: 1268- 1273.
  43. Cook MD, Allen JM, Pence BD, Wallig MA, gaskins HR, White BA, Woods JA. Exercise and gut immune function: evidence of alterations in colon immune cell homeostasis and microbiome characteristics with exercise training. *Immunol Cell Biol* 2016, 94: 158-163.
  44. Biragyn A, Ferrucci L. Gut dysbiosis: a potential link between increased cancer risk in ageing and inflammaging. *Lancet Oncol* 2018, 19: e295-e304.
  45. Man AWC, Li H, Xia N. Resveratrol and the Interaction between Gut Microbiota and Arterial Remodelling. *Nutrients*. 2020 Jan 1;12(1):119. doi: 10.3390/nu12010119. PMID: 31906281; PMCID: PMC7019510
  46. Myburgh KH. Polyphenol supplementation: benefits for exercise performance or oxidative stress? *Sports Med*. 2014 May;44 Suppl 1(Suppl 1):S57-70. doi: 10.1007/s40279-014-0151-4.
  47. Marttinen M, Ala-Jaakkola R, Laitila A, Lehtinen MJ. Gut Microbiota, Probiotics and Physical Performance in Athletes and Physically Active Individuals. *Nutrients*. 2020 Sep 25;12(10):2936. doi: 10.3390/nu12102936.
  48. Gubert C, Kong G, Renoir T, Hannan AJ. Exercise, diet and stress as modulators of gut microbiota: Implications for neurodegenerative diseases. *Neurobiol Dis*. 2020 Feb;134:104621. doi: 10.1016/j.nbd.2019.104621.
  49. Hughes RL, Holscher HD. Fueling Gut Microbes: A Review of the Interaction between Diet, Exercise, and the Gut Microbiota in Athletes. *Adv Nutr*. 2021 Dec 1;12(6):2190-2215. doi: 10.1093/advances/nmab077.
  50. Wegierska AE, Charitos IA, Topi S, Potenza MA, Montagnani M, Santacroce L. The Connection Between Physical Exercise and Gut Microbiota: Implications for Competitive Sports Athletes. *Sports Med*. 2022 Oct;52(10):2355-2369. doi: 10.1007/s40279-022-01696-x. Epub 2022 May 21. PMID: 35596883; PMCID: PMC9474385.
  51. Cataldi S, Bonavolontà V, Poli L, Clemente FM, De Candia M, Carvutto R, Silva AF, Badicu G, Greco G, Fischetti F. The Relationship between Physical Activity, Physical Exercise, and Human Gut Microbiota in Healthy and Unhealthy Subjects: A Systematic Review. *Biology (Basel)*. 2022 Mar 21;11(3):479. doi: 10.3390/biology11030479. PMID: 35336852; PMCID: PMC8945171.
  52. Campaniello D, Corbo MR, Sinigaglia M, Speranza B, Racioppo A, Altieri C, Bevilacqua A. How Diet and Physical Activity Modulate Gut Microbiota: Evidence, and Perspectives. *Nutrients*. 2022 Jun 14;14(12):2456. doi: 10.3390/nu14122456. PMID:

35745186; PMCID: PMC9227967.